

Cryptococcus Antigen with Reflex, Spinal Fluid

Test ID: LFACX; performed at Mayo Clinic Laboratories Florida.

Explanation:

As part of ongoing efforts to enhance testing efficiencies this test will become obsolete on October 8, 2026. The alternatives below are both recommended replacements for LFACX.

Recommended Alternative Test:

Cryptococcus Antigen Screen with Titer, Spinal Fluid

Test ID: CLFA; performed at Mayo Clinic Laboratories Florida.

Methodology:

Lateral Flow Assay (LFA)

Reference Values:

Negative

Reference values apply to all ages.

Specimen Requirements:

Container/Tube: Sterile vial

Specimen Volume: 1 mL

Collection Instructions: Submit specimen from collection vial 2 (preferred), 3, or 4.

Minimum Volume: 0.5 mL

Specimen Stability Information:

Specimen Type	Temperature	Time
CSF	Refrigerated (preferred)	14 days
	Frozen	14 days

CPT Code:

87899-Cryptococcus screen

87899-Cryptococcus titer (if appropriate)

Day(s) Performed: Monday through Sunday **Report Available:** 1 to 2 days

Recommended Alternative Test:

Fungal Culture, Routine

Test ID: FGEN

Methodology:

Conventional Agar Culture/Macroscopy/Microscopy/D2 and Internal Transcribed Spacer (ITS) rDNA Gene Sequencing/Matrix-Assisted Laser Desorption/Ionization Time-Of-Flight Mass Spectrometry (MALDI-TOF MS)
Dimorphic Pathogen Identification Confirmation: D2 and ITS rDNA Gene Sequencing/MALDI-TOF MS

Reference Values:

Negative

If positive, fungus will be identified.

Specimen Requirements:

Submit only 1 of the following specimens:

Preferred:

Specimen Type: Body fluid

Container/Tube: Sterile container

Specimen Volume: Entire collection

Specimen Type: Fresh tissue

Container/Tube: Sterile container

Specimen Volume: Pea size

Collection Instructions: Tissue should be placed in small amount of sterile saline or sterile water.

Acceptable:

Specimen Type: Bone marrow

Container/Tube: Sterile container, SPS/Isolator system, or green top (lithium or sodium heparin)

Specimen Volume: Entire collection

Specimen Type: Respiratory specimen

Container/Tube: Sterile container

Specimen Volume: Entire collection

Specimen Type: Urine

Container/Tube: Sterile container

Specimen Volume: 2 mL

Collection Instructions: Collect a random urine specimen.

Fresh tissue or body fluid are preferred over a swab specimen. Recovery of mycobacteria from swabs is generally very low yield. Only submit a swab specimen if tissue biopsy or fluid aspiration is not possible.

Specimen Type: Swab

Supplies: BD E-Swab (T853)

Sources: Dermal, ear, mouth, ocular, throat, or wound

Container/Tube: Sterile, screw-capped tube containing Liquid Amies Medium with flocked swab (eg, E-Swab)

Specimen Volume: 1 mL in swab container with swab

Collection Instructions:

1. Before collecting specimen, wipe away any excessive amount of secretion and discharge, if appropriate.
2. Obtain secretions or fluid from source with sterile flocked swab. **Paranasal sinus collections must use a nasopharyngeal flocked swab.**
3. Place flocked swab in sterile, screw-capped tube containing 1 mL of Liquid Amies Medium.
4. If smear and culture are requested or both a bacterial culture and fungal culture are requested, collect a second swab to maximize test sensitivity. Submit each swab in a separate sterile, screw-capped tube with 1 mL of Liquid Amies Medium.

Specimen Minimum Volume

Bone marrow or body fluid: 1 mL; Cerebrospinal fluid: 0.5 mL; Respiratory specimen: 1.5 mL; All other specimen types: See Specimen Required

Specimen Stability Information:

Specimen Type	Temperature	Time
Varies	Refrigerated (preferred)	7 days
	Ambient	7 days

CPT Code:

87102-Fungal culture, routine

87106-Id MALDI-TOF Mass Spec Yeast (if appropriate)

87107-Id MALDI-TOF Mass Spec Fungi (if appropriate)

87107-Fungal identification panel A (if appropriate)

87107-Fungal identification panel B (if appropriate)

87153-D2 fungal sequencing identification (if appropriate)

87153-ITS Fungal Sequencing (if appropriate)

87176-Tissue processing (if appropriate)

Day(s) Performed: Monday through Sunday **Report Available:** 24 to 35 days

Questions

Contact Betheney Wiles, Laboratory Resource Coordinator at 800-533-1710.